

Proceedings

Open Access

## Identifying single-nucleotide polymorphisms responsible for the linkage signal of rheumatoid arthritis on chromosome 6 by joint modeling of linkage and association

Wan-Yu Lin<sup>1</sup> and Daniel J Schaid\*<sup>2</sup>

Address: <sup>1</sup>Institute of Epidemiology, National Taiwan University, Floor 5, No. 17, Shuijhou Road, Zhongzheng District, Taipei 100, Taiwan and <sup>2</sup>Division of Biostatistics, Mayo Clinic, 200 First Street SW, Rochester, Minnesota 55905, USA

Email: Wan-Yu Lin - d92842006@ntu.edu.tw; Daniel J Schaid\* - schaid@mayo.edu

\* Corresponding author

from Genetic Analysis Workshop 15  
St. Pete Beach, Florida, USA. 11–15 November 2006

Published: 18 December 2007

BMC Proceedings 2007, 1(Suppl 1):S40

This article is available from: <http://www.biomedcentral.com/1753-6561/1/S1/S40>

© 2007 Lin and Schaid; licensee BioMed Central Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

This study evaluated the utility of unrelated controls and flanking markers when performing joint modeling of linkage and association by the LAMP software (version 0.0.6) [*Am J Hum Genet* 2005, 76:934–949; *Am J Hum Genet* 2006, 78:778–792]. Analyses were conducted on the simulated rheumatoid arthritis (RA) data in Genetic Analysis Workshop 15 (GAW15), using single-nucleotide polymorphisms (SNPs) on chromosome 6 over the 100 simulated replicates. We found that the LOD score for testing association in the presence of linkage dramatically increased when unrelated controls were added to affected sib pairs (ASPs), and that choosing a sufficient number of flanking markers is critical in order to distinguish between perfect linkage disequilibrium (which leads to the conclusion of a measured SNP explaining a linkage signal) and incomplete linkage disequilibrium (which leads to the conclusion of other undetected causal variants in a linkage region).

### Background

The DRB1 alleles located in the HLA region of chromosome 6 have been found to affect susceptibility to rheumatoid arthritis (RA), and linkage at HLA was confirmed by several studies [1-4]. However, because linkage measures the effect of a large chromosomal region, follow-up association studies are needed to identify a causative locus. Traditional association studies do not distinguish a causal gene and a gene with indirect effects through linkage disequilibrium (LD); rather, they test the presence of LD, instead of the presence of perfect LD. Among the measures of LD that have been proposed for two-locus

haplotype data, the two most common are  $|D'|$  and  $r^2$  [5,6]:  $|D'| = 1$  when the deviation of a haplotype frequency from randomly associated alleles attains its maximum value, given the marginal allele frequencies;  $r^2 = 1$  when two single-nucleotide polymorphisms (SNPs) are perfectly correlated, sometimes called "perfect LD". This can arise when two SNPs arose on the same branch of the genealogy and remain undisrupted by recombination. In contrast,  $r^2$  can have a value less than 1 when SNPs arose on different branches, or if an initially strong correlation has been disrupted by crossing over [7]. Here we distin-

guish between perfect LD ( $r^2 = 1$ ) and complete LD ( $|D'| = 1$ ).

To identify genes perfectly associated with disease, Li et al. [8] proposed a method to jointly model linkage and association, such that it can detect a marker's ability to explain a linkage signal, either partially or fully. They also discussed efficient study designs to test for association using sibship and unrelated individuals [9]. They found that when the disease is influenced by multiple genes, affected sib pairs (ASPs) provide more association information than singleton cases. Furthermore, a case-control study design can help to detect genes with small effects in the presence of genes with much larger effects.

To study the power of association tests in a linked region with different study designs, we used the SNP data on chromosome 6 for all 100 simulated replicates, and evaluated designs that either use only ASPs or combine ASPs with controls. All analyses were conducted with the software LAMP (version 0.0.6). We studied the simulated RA data with answers, to compare the LOD scores provided by LAMP under the two designs.

**Methods**

**Phenotype, genotypes, and map**

Using RA affection status as a binary trait, there were 1500 families with one ASP and their parents, all genotyped on 674 SNPs along chromosome 6. Additionally, 2000 unrelated controls were included in each replicate. There were no missing data or genotype errors. For the LAMP analyses, we specified the lifetime prevalence of RA to be 0.0107 as stated in the introduction of the simulated RA data. The sex-averaged map locations (in Haldane centimorgans) were used for the maps.

**Four analysis models**

The software LAMP was used to fit four models by maximum likelihood: 1) a base model (BM) for no linkage and no association,  $L(\theta = \frac{1}{2}, r^2 = 0)$ , where  $\theta$  is the recombination fraction and  $r^2$  is the measure of LD. LAMP esti-

mates marker allele frequencies with the assumption of Hardy-Weinberg equilibrium, and there is only one fitted parameter for each SNP; 2) a linkage equilibrium model (LE) for linkage without association,  $L(\theta = 0, r^2 = 0)$ . LAMP estimates disease and marker allele frequencies, as well as the penetrances for disease genotypes. The estimation of these parameters is constrained by the assumed disease prevalence; 3) a general model (GM) for linkage with any level of association,  $L(\theta = 0, 0 < r^2 < 1)$ . This most general model estimates three marker-disease haplotype frequencies and the penetrances for disease genotypes; 4) a linkage disequilibrium model (LD) for linkage with complete association,  $L(\theta = 0, r^2 = 1)$ . For this model, one of the marker alleles is assumed to directly affect disease susceptibility. The marker allele frequency and the penetrances for disease genotypes are estimated. The four models are summarized in Table 1.

**Linkage and association tests**

The four models were used to create three likelihood ratio tests, and hence LOD scores: 1) a test for linkage, 2) a test for association in the presence of linkage, and 3) a test for other linked variants. In contrast to traditional parametric linkage analyses that specify a genetic disease model, LAMP calculates parametric maximized LOD scores by maximizing over all assumed model parameters. The method allows LD between the measured candidate SNP and an unobserved disease allele, but assumes linkage equilibrium between the flanking markers and the measured candidate SNP [8]. Note that this assumption may not hold for high-density SNP data. For the 674 SNPs on chromosome 6, there were 62 marker clusters that had large within-block LD, identified by setting the  $r^2$  threshold to 0.4 in Merlin (version 1.0.1, with a command "merlin -d datfile -p pedfile -m mapfile --npl --grid 1 --rsq 0.4 --cfreq") [10,11]. To avoid problems of LD between the candidate SNP and the flanking markers, we eliminated SNPs that had large LD values by selecting a tag-SNP from each cluster. In each marker cluster, we kept the SNP with the highest marker heterozygosity (calculated by pedstats,

**Table 1: Four models compared in the three likelihood-ratio tests**

Model <sup>a</sup>	Likelihood	d.f.	Parameters <sup>b</sup>	LR tests
(1) BM	$L(\theta = \frac{1}{2}, r^2 = 0)$	1	$p_A$	
(2) LE	$L(\theta = 0, r^2 = 0)$	4	$p_A, p_D, f_{DD}, f_{Dd}, f_{dd}$	(2) vs. (1): test for linkage
(3) GM	$L(\theta = 0, 0 < r^2 < 1)$	5	$p_{DA}, p_{Da}, p_{dA}, f_{DD}, f_{Dd}, f_{dd}$	(3) vs. (2): test for association in the presence of linkage
(4) LD	$L(\theta = 0, r^2 = 1)$	3	$p_A, f_{DD}, f_{Dd}, f_{dd}$	(3) vs. (4): test for other linked variants

<sup>a</sup>BM, base model; LE, linkage equilibrium; GM, general model; LD, linkage disequilibrium.

<sup>b</sup> $p_A$  = marker allele frequency;  $p_D$  = disease allele frequency;  $f_g = P(\text{affected} | g)$ ,  $g \in \{DD, Dd, dd\}$ ;  $p_{DA}, p_{Da}, p_{dA}$  are the marker-disease haplotype frequencies.

version 0.6.3), and deleted other SNPs in the same cluster. There were 581 tag-SNPs after removing those with large LD.

**Two study designs**

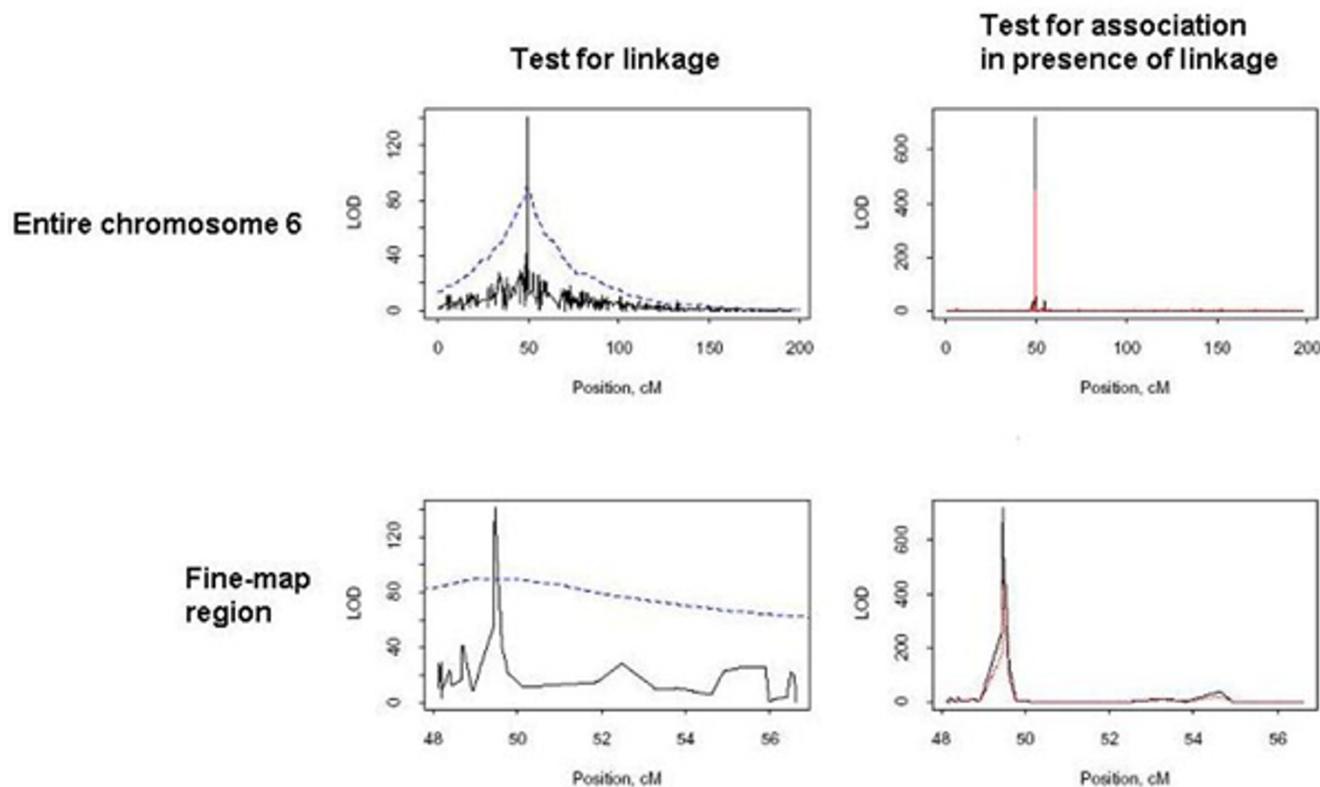
We considered two study designs: 1500 ASPs alone (ASPs), and 1500 ASPs with unrelated 2000 controls (ASPs+controls). The combined data should be more powerful to detect association, but not linkage. Our aims were to evaluate the increase in LOD scores provided by controls for the test for association (in the presence of linkage), and to evaluate the benefit of flanking markers for the test for other linked variants.

**Results**

**Contribution of unrelated controls to LOD scores**

Results from the simulated replicates were very consistent, as summarized in Table 2, including the mean and range of LOD scores at SNP6\_153 and SNP6\_162 across the 100 replicates. These two SNPs were selected because they

caused peaks for the test of association in the presence of linkage (see the second column of Figure 1). The peak linkage was at SNP6\_153 (49.461 cM) in 75 out of 100 replicates, and at SNP6\_154 (49.466 cM) in the other 25 replicates. Note that the two study designs resulted in the same LOD scores for linkage, since unrelated controls provided no information for linkage. The LOD score for association in the presence of linkage showed an extremely large value at SNP6\_153 in all replicates for both study designs. Another signal was detected at SNP6\_162 (54.625 cM). From Table 2, we can see that controls increased the LOD scores for the tests for association in the presence of linkage by approximately 60%. Furthermore, for SNP6\_153, adding controls significantly increased the LOD score for the test for other linked variants, suggesting that SNP6\_153 is not in perfect LD with a disease allele, and hence, that there may be other associated alleles in the region. This result agrees with the simulation scenario. In the simulated data, none of the 674



**Figure 1**  
**LOD scores for test of linkage and test of association in the presence of linkage.** Simulated Replicate 45, representative of most 100 replicates, was randomly selected to illustrate key features. For the tests of linkage (left column), the black solid lines are the LOD scores obtained without the flanking markers, while the blue broken lines are the LOD scores obtained with a grid of flanking markers in the surrounding 30-cM region. For the tests for association in the presence of linkage (right column), the black solid lines are the LOD scores for ASPs+controls, while the red broken lines are the LOD scores for ASPs.

**Table 2: Mean LOD scores for the three tests summarized from 100 replicates**

	Test for linkage	Test for association in the presence of linkage	Test for other linked variants	
Models compared	LE vs. BM	GM vs. LE	GM vs. LD	
d.f.	4 - 1 = 3	5 - 4 = 1	5 - 3 = 2	
SNP6_I53			With flanking markers	Without flanking markers
ASPs+controls		705.3 (range: 653.1–763.0)	7.8 (range: 2.2–17.9) (power <sup>a</sup> : 100%)	6.8 (range: 1.8–15.5) (power: 100%)
ASPs	132.3 (range: 105.9–152.9)	441.9 (range: 401.3–485.8)	4.1 (range: 0.8–12.6) (power: 94%)	1.7 (range: 0.0–5.3) (power: 53%)
SNP6_I62				
ASPs+controls		44.4 (range: 30.0–62.2)	59.1 (range: 39.5–81.1) (power: 100%)	2.0 (range: 0.0–6.3) (power: 63%)
ASPs	9.1 (range: 3.4–15.8)	27.8 (range: 14.8–41.3)	58.9 (range: 39.7–81.0) (power: 100%)	1.8 (range: 0.0–6.1) (power: 55%)

<sup>a</sup>Power based on significance level of 0.05 and asymptotic chi-square distribution.

SNPs on chromosome 6 was in perfect LD ( $r^2 = 1$ ) with the causal locus.

#### Contribution of flanking markers to LOD scores

In addition to the impact of unrelated controls, flanking markers can have a large impact on LOD scores. Flanking markers smooth the LOD scores of the test for linkage, as we can see from the first column of Figure 1. Moreover, they are necessary to distinguish between perfect LD versus other linked variants, because this test relies on the evidence for linkage in the chromosomal region. To evaluate the impact of flanking markers, we used the SNPs located from 40 to 60 cM as the flanking markers, although only tag-SNPs were used. This region was highlighted because it presented a strong linkage signal, as we can see from Figure 1 (blue line in the upper-left plot). The black line in the first column of Figure 1 illustrates that the LOD scores for linkage in the linked region were drastically reduced when flanking markers were not used. Table 2 illustrates that the LOD scores for other linked variants were much too small when flanking markers were not used, giving the misleading conclusion that these two SNPs are in perfect LD with a disease causing allele.

#### Discussion and conclusion

The joint modeling of linkage and association proposed by Li et al. [8] uses genotype information contributed by both the candidate SNP and the flanking markers, which they claimed outperforms the method of Göring and Terwilliger [12]. Their method identifies association in the presence of linkage, and can sometimes distinguish between perfect LD (leading to the conclusion of a causative SNP) and incomplete LD (leading to the conclusion

of other causal variants in the region). A limitation is the assumption of linkage equilibrium between the candidate SNP and the flanking markers. This assumption is hard to meet with high-density SNP data. One way to eliminate LD among markers is to select tag-SNPs from each marker cluster. Further research to improve use of markers with LD would be worthwhile, recognizing that this might increase computational intensity.

In our analyses, we used sex-averaged maps without considering the sex differences in genetic map distances. The results of testing for linkage, testing for association in the presence of linkage, and testing for other linked variants were consistent with the simulation answers. However, ignoring sex differences in genetic maps may not lead to accurate inferences when genotypes for only one parent of each ASP are available [13]. A drawback of LAMP is that it is computationally demanding to search for the maximum likelihood estimates, particularly when pedigree data and microsatellite markers are used. To increase power to test for association in the presence of linkage without much computational intensity, combining ASPs with unrelated controls is a worthwhile design. Finally, our results emphasize the importance of choosing a sufficient number of flanking markers to distinguish between perfect LD and incomplete LD.

#### Competing interests

The author(s) declare that they have no competing interests.

#### Acknowledgements

We are grateful for the helpful comments from the editors and the anonymous reviewers that improved this work. This work was supported by the

U.S. Public Health Service, National Institutes of Health, contract grant number GM67768 (DJS) and the National Science Council of Taiwan NSC 095-2917-I-002-012 (W-YL). We thank members of GAW15 presentation group for helpful discussions.

This article has been published as part of *BMC Proceedings* Volume 1 Supplement 1, 2007: Genetic Analysis Workshop 15: Gene Expression Analysis and Approaches to Detecting Multiple Functional Loci. The full contents of the supplement are available online at <http://www.biomedcentral.com/1753-6561/1?issue=S1>.

## References

1. Etzel CJ, Chen WV, Shepard N, Jawaheer D, Cornelis F, Seldin MF, Gregersen PK, Amos CI, For the North American Rheumatoid Arthritis Consortium: **Genome-wide meta-analysis for rheumatoid arthritis**. *Hum Genet* 2006, **119**:634-641.
2. Jawaheer D, Lum RF, Amos CI, Gregersen PK, Criswell LA: **Clustering of disease features within 512 multicase rheumatoid arthritis families**. *Arthritis Rheum* 2004, **50**:736-741.
3. Jawaheer D, Seldin MF, Amos CI, Chen WV, Shigeta R, Etzel C, Damle A, Xiao X, Chen D, Lum RF, Monteiro J, Kern M, Criswell LA, Albani S, Nelson JL, Clegg DO, Pope R, Schroeder HW Jr, Bridges SL Jr, Pisetsky DS, Ward R, Kastner DL, Wilder RL, Pincus T, Callahan LF, Flemming D, Wener MH, Gregersen PK, North American Rheumatoid Arthritis Consortium: **Screening the genome for rheumatoid arthritis susceptibility genes: a replication study and combined analysis of 512 multicase families**. *Arthritis Rheum* 2003, **48**:906-916.
4. Osorio Y, Fortea J, Bukulmez H, Petit-Teixeira E, Michou L, Pierlot C, Cailleau-Moindraut S, Lemaire I, Lasbleiz S, Alibert O, Quillet P, Bardin T, Prum B, Olson JM, Cornelis F: **Dense genome-wide linkage analysis of rheumatoid arthritis, including covariates**. *Arthritis Rheum* 2004, **50**:2757-2765.
5. Devlin B, Risch N: **A comparison of linkage disequilibrium measures for fine-scale mapping**. *Genomics* 1995, **29**:311-322.
6. Balding DJ: **A tutorial on statistical methods for population association studies**. *Nature Rev Genet* 2006, **7**:781-791.
7. The International HapMap Consortium: **A haplotype map of the human genome**. *Nature* 2005, **437**:1299-1320.
8. Li M, Boehnke M, Abecasis GR: **Joint modeling of linkage and association: identifying SNPs responsible for a linkage signal**. *Am J Hum Genet* 2005, **76**:934-949.
9. Li M, Boehnke M, Abecasis GR: **Efficient study designs for test of genetic association using sibship data and unrelated cases and controls**. *Am J Hum Genet* 2006, **78**:778-792.
10. Abecasis GR, Cherny SS, Cookson WO, Cardon LR: **Merlin: rapid analysis of dense genetic maps using sparse gene flow trees**. *Nat Genet* 2002, **30**:97-101.
11. Abecasis GR, Wigginton JE: **Handling marker-marker linkage disequilibrium: pedigree analysis with clustered markers**. *Am J Hum Genet* 2005, **77**:754-767.
12. Göring HHH, Terwilliger JD: **Linkage analysis in the presence of errors. IV: Joint pseudomarker analysis of linkage and/or linkage disequilibrium on a mixture of pedigrees and singletons when the mode of inheritance cannot be accurately specified**. *Am J Hum Genet* 2000, **66**:1310-1327.
13. Fingerlin TE, Abecasis GR, Boehnke M: **Using sex-averaged genetic maps in multipoint linkage analysis when identity-by-descent status is incompletely known**. *Genet Epidemiol* 2006, **30**:1384-1396.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

